

## The best strategy of blood transfusion in patients with Rh<sub>null</sub> syndrome

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### ABSTRACT

Rh<sub>null</sub> syndrome is a very rare disease. Patients with this syndrome present with negative serological Rh typing of E, e, C, c, and D antigens. Only one study has previously discussed Rh<sub>null</sub> syndrome in Chinese individuals. We experienced two patients with Rh<sub>null</sub> syndrome in China, Rh genotypes being CcDEe in the first patient and CCDee in the second patient. The first patient was a pregnant woman (gravida 2, para 1) with a negative red blood cell (RBC) antibody screen test. The second patient was a middle-aged man, transfused with ccdee, ccdEe, and ccdee RBC products, the pre-transfusion specimen was negative and post-transfusion specimen was anti-c,e, respectively. The hemoglobin level continued to increase in the second patient after being transfused with ccdEe RBC products. In the first patient, the result of the antibody screen test was still negative after artificial abortion. In patients with Rh<sub>null</sub> syndrome, RBC products that have the same Rh genotype as the patient can be safely transfused.

**Keywords:** Rh<sub>null</sub> syndrome, Rh associated glycoprotein, transfusion strategy

### INTRODUCTION

Individuals with Rh<sub>null</sub> syndrome are negative for RhD and RhCE genotypes<sup>[1]</sup>, which is caused by mutations of the RhCE/RhD gene (Rh<sub>null</sub> amorph type) and mutations of the Rh-associated glycoprotein (RhAG) (the regular Rh<sub>null</sub> type)<sup>[2]</sup>. Previous studies have demonstrated that individuals with Rh<sub>null</sub> syndrome usually have the anti-Rh29 antibody, making transfusion a challenge<sup>[3,4]</sup>. We described one patient with Rh<sub>null</sub> syndrome, who was immune to different Rh types of red blood cell (RBC) products and presented with pregnancy, and another patient transfused with different Rh types of RBC products. This report

aimed to suggest the best transfusion strategy for patients with the rare Rh<sub>null</sub> syndrome.

### METHODS

#### Antibody screen test, identification and phenotyping

Screen and panel cells used at the Blood Group Laboratory (BGL) of the Beijing Red Cross Blood Center (BRCBC) cover antigen systems, including Rh, MNS, Duffy, Kidd, Kell, Lewis, Pl, Xg, Lutheran, Dia and Mur (Bio-Rad Laboratories GmbH, Germany; Jiangsu LiBio Biotech, China), which have a relatively high prevalence in Asia. Detection methods include conventional tube techniques in room temperature and gel column tests (Bio-Rad Laboratories GmbH, Germany; Jiangsu LiBio Biotech, China). The Rh phenotype was confirmed in column agglutina-

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tion testing (Bio–Rad Laboratories GmbHMunchen, Germany; Jiangsu LiBio Biotech, China). Other procedures, such as sample reception, report documentation, and report writing were all based on regulations and standard procedures specified by the BGL of the BRCBC.

### Genotyping and molecular analysis

Polymerase chain reaction with sequence–specific primers was performed for the Rh blood group genotyping by using commercial kit (Multi–erythrocyte antigen genotype, Jiangsu LiBio Biotech, China). The sequence primers and amplification protocol for RhD exons 1–10 and RhCE exons 1–10 were designed using the Sanger sequence method by a commercial kidd gene kit (RhD and RhCE whole exons sequence, Jiangsu LiBio Biotech, China). Sequencing of PCR purified products was done by Sangon Biotech (Beijing, China) and the results were analyzed using sequence analysis software (Geneious R9, New Zealand). The RhD and RhCE allele sequence (GenBank ID: No. 6006 and 6007) template was used as reference to analyze and mark the mutations.

## RESULTS

### Case 1

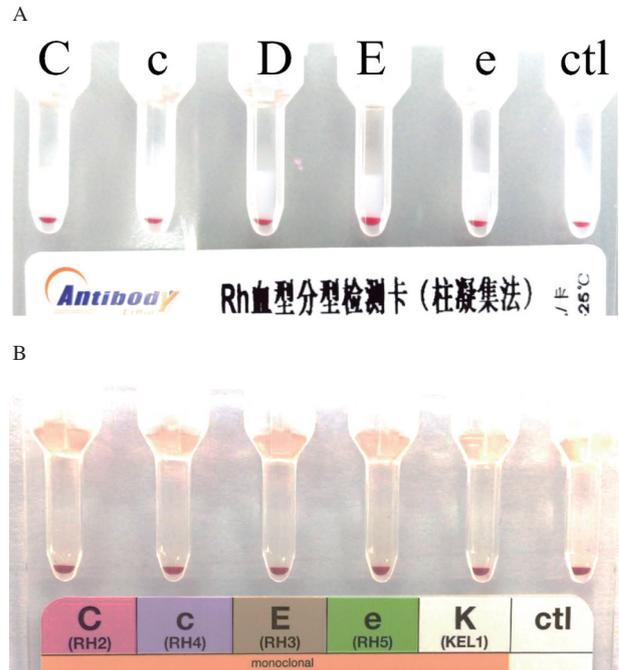
A 21–year–old Chinese woman, gravida 2, para1, presented at 18 weeks gestation, with a history of chronic anemia. She had been diagnosed as being Rh serological results showing negative during an antenatal examination, and phenotyping results demonstrated that she was negative for RhCE and RhD antigens (**Fig.1 A**, **Fig.2 A**). The result of the antibody screen test was negative, and the Rh genotype was CcDEe. We examined the Rh genotype by sequencing the exons of RhD and RhCE, and the results showed CcDEe.

### Case 2

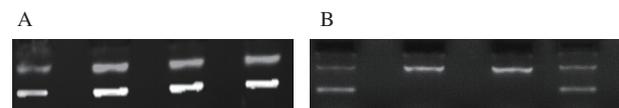
A 53–year–old man was referred for mitral and aortic valve replacement to treat rheumatic heart disease in 2002. In 2009, a Doppler echocardiogram showed tricuspid regurgitation, and drug treatment was prescribed with continuous follow–up until March 2017. Then he underwent tricuspid valve replacement with mechanical valves and was hospitalized in April 2017. The routine preoperative antibody screen test result was negative, and the serological blood type was A and negative for D (**Fig.1 B**, **Fig.2 B**). Genotyping was conducted, and all RhCE and RhD antigens were negative; thus, we initially planned to transfuse RhD–negative RBC products. Since the

hemoglobin (Hb) level was 119 g/L, we obtained autologous whole blood that was stored preoperatively. EPO and iron supplements had been given since 2 weeks before phlebotomy was performed.

The patient received auto–transfusion with autologous scavenged RBCs(1,300 mL) intraoperatively.



**Fig.1. The serological Rh typing for the  $Rh_{null}$  syndrome.** A showed C, c, D, E, e phenotyping negative in first patient with  $Rh_{null}$  syndrome. B showed D, C, c, E, e phenotyping negative in second patient with  $Rh_{null}$  syndrome and control result showed negative.



**Fig.2. The Rh genotyping for the  $Rh_{null}$  syndrome.** The upper white band human growth hormone receptor(hGHR) gene fragment as internal control and lower white band index positive antigen for C, c, E, or e(from left to right). A showed the first patient was CcDEe in genotyping; B showed second patient was CCDEe in genotyping.

After tricuspid valve replacement was finished, autologous pre–stored whole blood ( $2 \times 200$  mL collected from 7 and 14 days preoperatively) was transfused. In addition, suspended ccdee RBCs (2 units) were given on day 1, and fresh–frozen ccdee RBCs (2 units) were transfused on day 2. Later, suspended ccdEe RBCs (2 units) were transfused again on day 8, and fresh–frozen ccdee RBCs (1.5 units) were transfused on day 11. The antibody screen test result was negative at every pre–transfusion examination until day 13 postoperatively. The antibody screen test identified the anti–f (anti–c,e) in the patient’s plasma on day 13, and the direct agglutination test showed a weak positive

result. Other data, including the Hb level, are shown in **Table 1**. The Rh genotyping and sequence of Rh exons confirmed the CCDee genotype, so we decided to use CCDee RBC products instead of serologically

RhD–negative RBC products. Since the Rh genotype of blood products corresponded to the patient’s genotype, the Hb level continued to increase from 64 g/L on day 11 to 82 g/L on day 18.

**Table 1 The Hb level, unexpected antibody screen test and transfusion record of case 2**

| Date                   | Hb(g/L)  | Blood products                                 | Blood type      | Unexpected antibody screen test |
|------------------------|----------|------------------------------------------------|-----------------|---------------------------------|
| 2 weeks before surgery | 119      | draw autologous pre–stored whole blood, 200 mL |                 |                                 |
|                        | 115, 118 | draw autologous pre–stored whole blood, 200 mL |                 |                                 |
| Surgery, Day 0         | 97       | autologous scavenged whole blood, 400 mL       | patient’s blood | Negative                        |
|                        |          | washed autologous scavenged red cell, 400 mL   | patient’s blood |                                 |
|                        |          | washed autologous scavenged red cell, 500 mL   | patient’s blood |                                 |
|                        |          | autologous pre–stored whole blood, 400 mL      | patient’s blood |                                 |
| Day 1                  | 82, 79   | red cells suspension, 2 units                  | A, ccdee        | Negative                        |
| Day 2                  | 78, 70   | frozen red blood cells, 2 unit                 | A, ccdee        | Negative                        |
| Day 8                  | 69       | red cells suspension, 2 units                  | A, ccdEe        | Negative                        |
| Day 11                 | 64       | frozen red blood cells, 1.5 unit               | A, ccdee        | Negative                        |
| Day 13                 |          |                                                |                 | anti–f                          |
| Day 16                 |          | red cells suspension, 2 units                  | A, CCDee        |                                 |
| Day 17                 |          | red cells suspension, 2 units                  | A, CCDee        |                                 |
| Day18                  | 82       |                                                |                 |                                 |
| Day21                  | 77       |                                                |                 |                                 |
| Day24                  | 87       |                                                |                 |                                 |

## DISCUSSION

In patients with the Rh<sub>null</sub> phenotype, no Rh antigens are found on the surface of the RBC membrane, and the morphologic changes of the RBCs include spherocytes, stomatocytes, and an isocytosis<sup>[5,6]</sup>. Rh<sub>null</sub> syndrome induces abnormal organization of RBC membrane phospholipids, increased activity of Na<sup>+</sup>K<sup>+</sup> pump and subsequential imbalance of water and ions in the cells<sup>[7]</sup>. Rh<sub>null</sub> cells usually reduce the survival period, lower resistance to osmolality, increase the reticulocyte count, and cause varying degrees of hemolysis because of the abnormal morphology and cell membrane contents. Patients with Rh<sub>null</sub> syndrome require transfusion during operations or instances of massive bleeding. Therefore, the strategy of blood transfusion is an important issue.

Several immunized patients with Rh<sub>null</sub> syndrome with anti–Rh29 (allo–antibodies reactive with total Rh antigens) and other immunized patients with the Rh<sub>null</sub> syndrome have been reported to have anti–Hr0 (allo–antibodies reactive with E, e, C, and c antigens) or anti–e<sup>[3,4,8,9]</sup>. In our study, we observed a pregnant woman with a history of artificial abortion and a cardiac patient immunized to different Rh types of RBC products. The second patient produced anti–c,e by ccdee RBCs, and was transfused with CCDee RBCs, which was the same genotype as his suspended RBCs; this was effective for increasing the Hb level. Several patients with Rh<sub>null</sub> syndrome with unexpected antibodies have undergone repeated blood exchange transfusion successfully, including the pregnant wom-

an and cardiac patient in our study.

Although the D antigen frequently occurs (99.6%) in China, allo–antibodies against Rh antigens are still the most frequent challenge of blood transfusions in China<sup>[10]</sup>. Rh phenotyping is routinely conducted at several hospitals in China, and prophylaxis in patients to ensure transfusion matching could effectively reduce the occurrence of the unexpected antibody. For some patients, e.g. those with auto–control positive samples, a Ce deficiency, and Rh<sub>null</sub> syndrome, genotyping of RBC groups is an advanced tool for solving the challenge of transfusion.

We observed two patients with Rh<sub>null</sub> syndrome, immunized to different Rh types of RBC products. We also demonstrated unexpected results of the antibody screening test (i.e. a negative result can become positive for anti–c,e after ccdee RBC transfusion). Rh genotyping matched RBCs may be the best strategy of blood transfusion in patients with Rh<sub>null</sub> syndrome to use.

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